

Microfluidic PCR in diagnostic microbiology - an overview.

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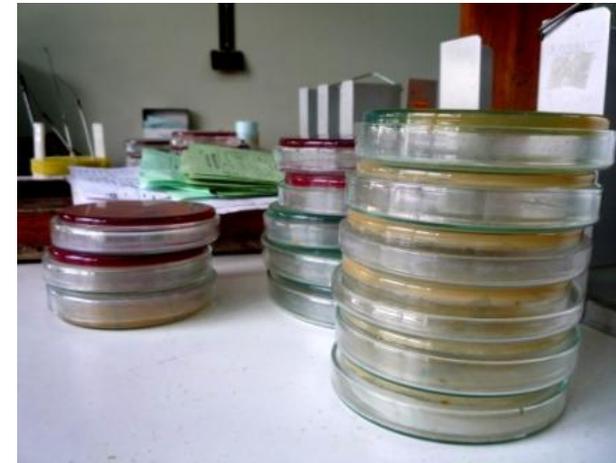
Supervisor: Prof Margaret Ip

Outline

- Laboratory diagnosis of infectious diseases
- Microfluidic PCR
- Applications of microfluidic PCR in diagnostic microbiology
- Issues and future

Diagnostic microbiology

- Laboratory diagnosis of infectious diseases
- Why? - aetiological diagnosis, tailor made treatment, infection control, epidemiology and prevention
- How? – Conventionally, by culture based methods and serology
- Time consuming, requires equipped laboratories, training, man power
- Quest for better tests –
Molecular methods



PCR in Diagnostic Microbiology

- “Rapid”, Accurate, Better sensitivity
- “Time consuming” – high thermal mass
- Reagents –expensive, requires specific storage conditions
- Expensive bulky equipment
- Specific laboratory designs – (contamination)
- Trained personnel



Centralized laboratories
? Point of care diagnosis

Microfluidic systems

- Small volumes of fluids are manipulated precisely in platforms fabricated with micro pumps, valves, etc

Micro total analysis systems (μ TAS)/

Lab-on-a-chip (LOC)

- Applications in diagnostic microbiology

Lateral flow devices for rapid diagnosis

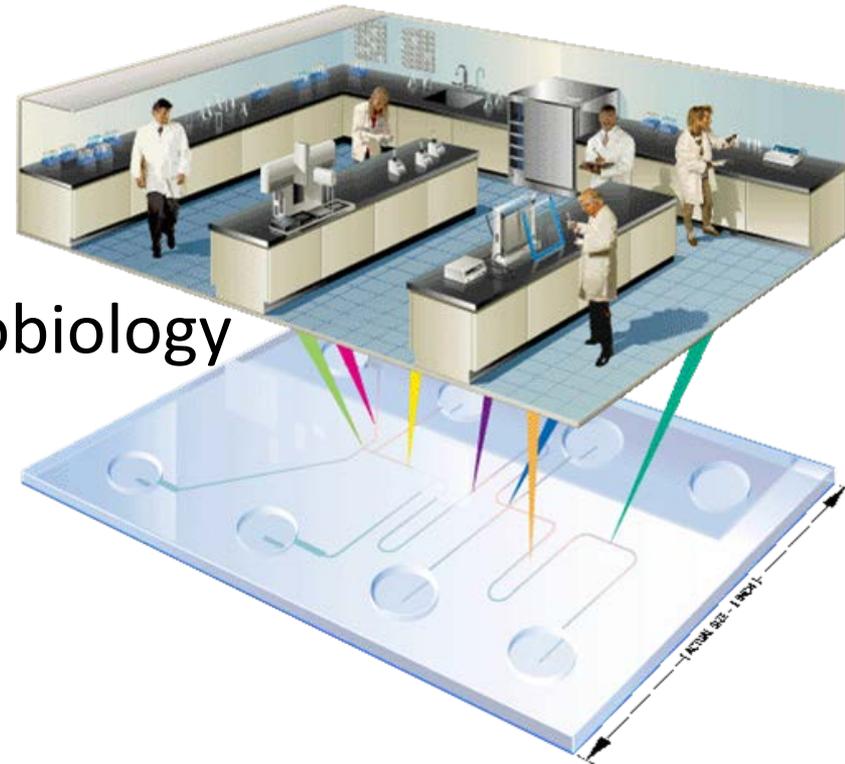
Culture

ABST

PCR

Microarray

Sequencing



©<http://www.gene-quantification.de/lab-on-chip.html>

Microfluidic PCR

First described in 1993 by Northrup *et al*

Potential advantages over conventional PCR

Faster speed

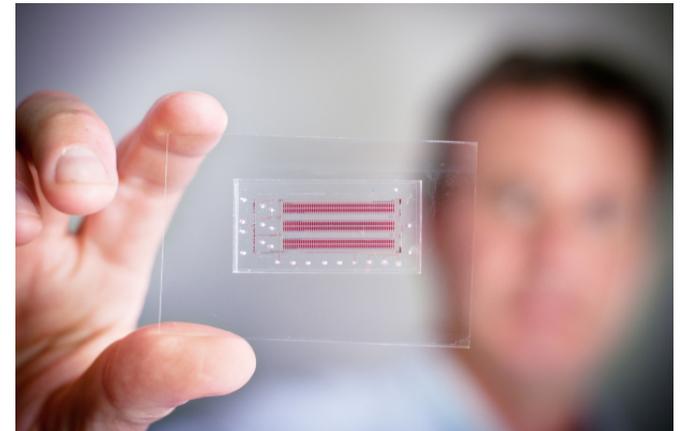
Less reagent usage

Automation

Complete integration

High throughput

Portability



(Credit: University of British Columbia) http://news.cnet.com/8301-27083_3-20083814-247/new-lab-on-a-chip-genetic-analysis-resembles-pinball/

Microfluidic PCR

- Reaction volumes

0.45nl – 50 μ l

- Reagents

Droplet based technology Vs dry reagent

- Heating methods

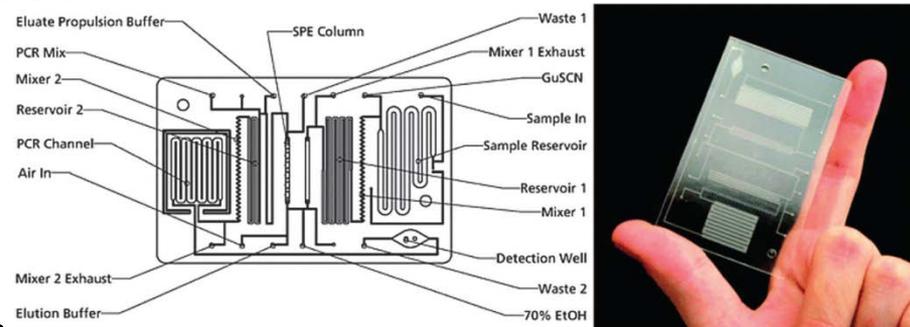
Contact Vs non contact

- Heating and cooling rates

175 $^{\circ}$ C/s – 2 $^{\circ}$ C/s

- Materials

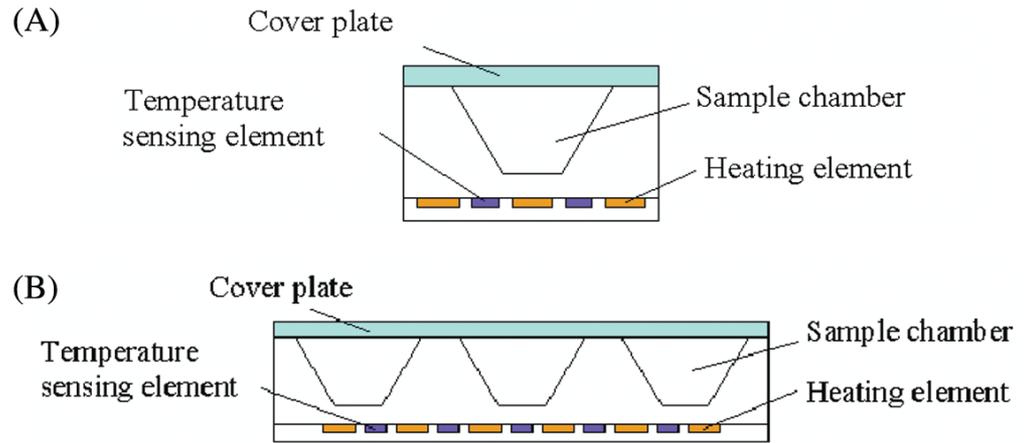
Glass, silicon, polymers



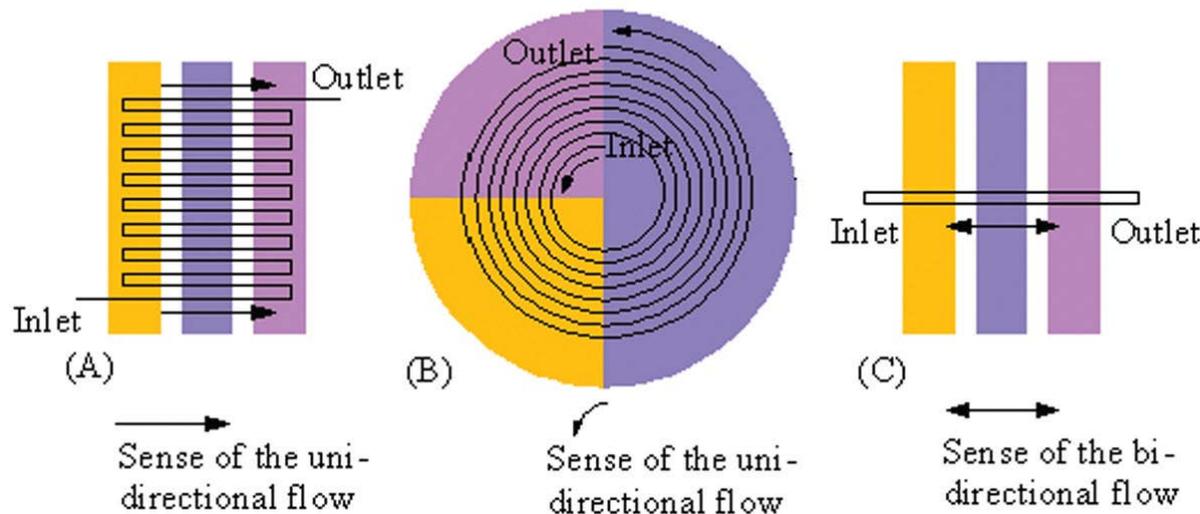
Microfluidic PCR systems

Chips -

- Stationary chamber Vs Continuous flow



- Denaturation temperature
- Extension temperature
- Annealing temperature



©Zhang C, Xing D. Miniaturized PCR chips for nucleic acid amplification and analysis: latest advances and future trends. *Nucleic Acids Res.* 2007;35(13):4223-37.

Pre PCR processing

- Low levels of organisms responsible and complex nature of samples (inhibitors, other organisms)
- Options available
 - Off chip samples processing – conventional extraction methods
 - On chip samples processing – complicates fabrication
separation, lysis, concentration
 - Use of unprocessed samples –overcoming inhibitions by using special Taq polymerases

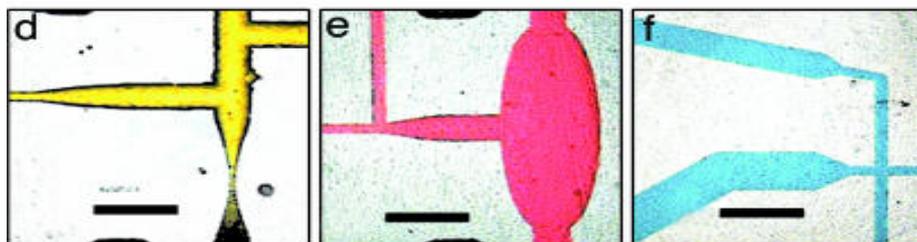
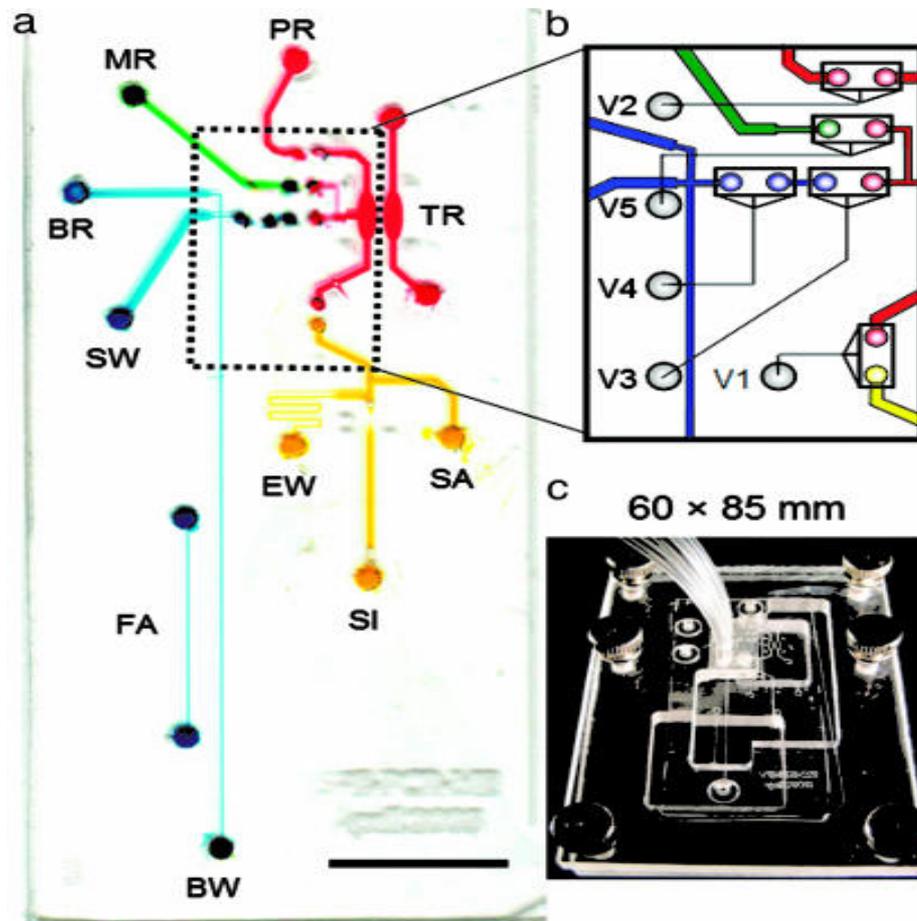
Post PCR applications

- Options available
 - Off chip Vs On chip
 - Capillary electrophoresis
 - Lateral flow techniques
 - DNA hybridization
 - Real time methods
 - Electrochemical sensing

A fully integrated microfluidic genetic analysis system with sample-in-answer-out capability.

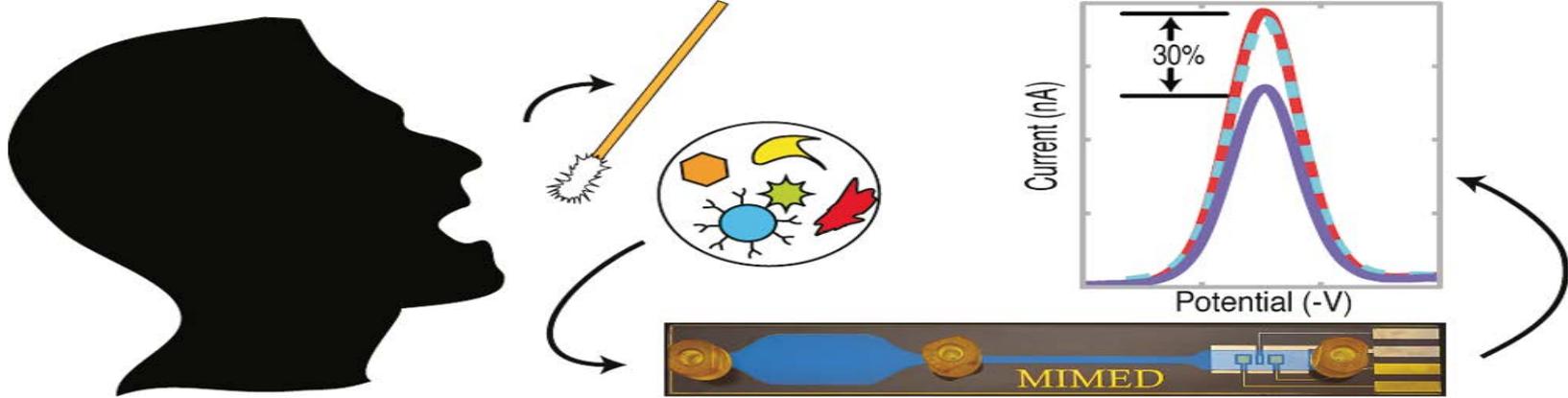
Easley CJ, Karlinsey JM, Bienvenue JM, Legendre LA, Roper MG, Feldman SH, Hughes MA, Hewlett EL, Merkel TJ, Ferrance JP, Landers JP.

Department of Chemistry, University of Virginia, Charlottesville, VA 22904, USA.



Applications in diagnostic microbiology

- Numerous papers – Dengue, Hep B, MRSA, SARS corona etc
- Initial work concentrates on chip thermal cycling
- Integrated methods Vs isolated use of one component
- Direct detection from patient samples Vs characterization of isolates
- Majority of studies conducted in research settings with spiked samples to represent clinical samples



Ferguson BS *et al.* Genetic analysis of H1N1 influenza virus from throat swab samples in a microfluidic system for point-of-care diagnostics. *J Am Chem Soc.* 2011 Jun 15;133(23):9129-35.

- Throat swab + antibody-coated magnetic beads+ RNA stabilizer in a tube
- Pumped into the device at 60 ml/h
- RNA extraction
- RT PCR Mix injected
- RT PCR
- ssDNA generation
- Detection by hybridization
- Chip dimensions - 1 x 6 cm
- Sample – result time – 3.5 hours (150 min for RTPCR)
- Detection limit ≈ 10 TCID₅₀

Microfluidic Platform versus Conventional Real-time PCR for the Detection of *Mycoplasma pneumoniae* in Respiratory Specimens

Elizabeth Wulff-Burchfield¹, Wiley A. Schell², Allen E. Eckhardt³, Michael G. Pollack³, Zhishan Hua³, Jeremy L. Rouse³, Vamsee K. Pamula³, Vijay Srinivasan³, Jonathan L. Benton², Barbara D. Alexander², David A. Wilfret⁴, Monica Kraft⁵, Charles Cairns⁶, John R. Perfect², and Thomas G. Mitchell^{7,*}

Comparison of real-time PCR results of acute patient NPWs on conventional and microfluidic real-time PCR platforms

		Conventional real-time PCR	
		Positive	Negative
Microfluidic real-time PCR	Positive	2	0
	Negative	1	56

Commercial applications

- At lab on a cartridge level
- Expensive
- Needs bulky equipment, uninterrupted power supply
- Eg – WHO endorsed Xpert MTB/RIF assay
 - Other systems by Cepheid ,
 - Microfluidic systems,
 - Fluidigm etc



Issues

- Integration and fabrication
- Adsorption of reagents and samples by surfaces and evaporation
- Inhibition of PCR by certain products used in fabrication of devices
- Validation for diagnostic use

Future

- Multiplexing
- Not just pathogen detection, but detection of virulence and antibiotic resistance markers
- Organism sensing and detections of biomarkers for infection together
- Point of care application in resource limited setting

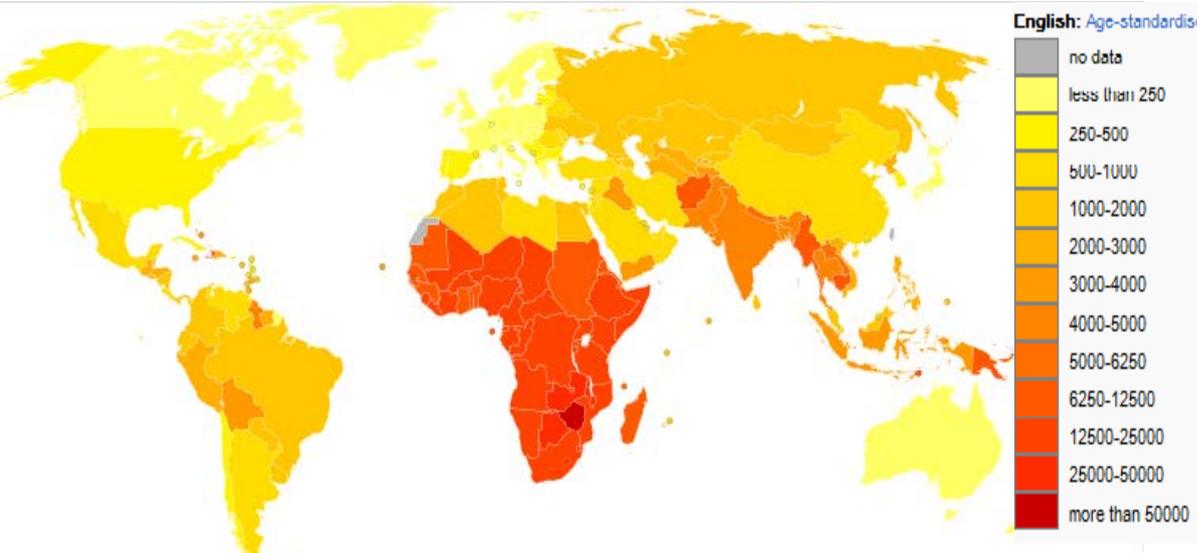
Distribution of commercial ventures in microfluidic technologies



<http://healthcareconsiderations.blogspot.com/2010/08/navigating-paths-and-protocols-of.html>

<http://fluidicmems.com/list-of-microfluidics-lab-on-a-chip-and-biomems-companies/>

Disability adjusted life years from infections and parasitic diseases (compiled with WHO 2004 data)



http://en.wikipedia.org/wiki/File:Infectious_and_parasitic_diseases_world_map_-_DALY_-_WHO2004.sva#filelinks

Lab on a chip or chip in a lab?

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Thank You!